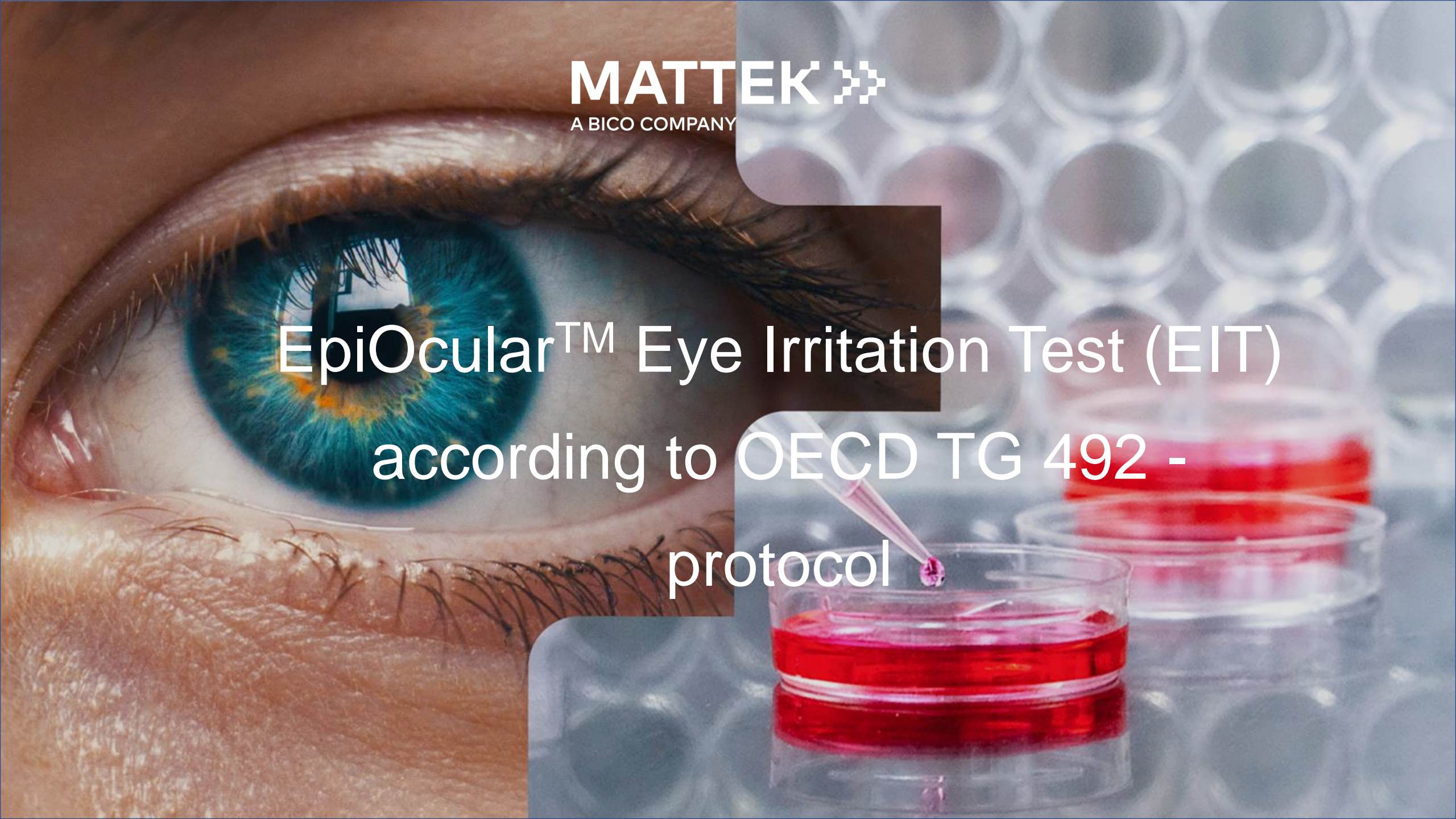


MATTEK >>
A BICO COMPANY

EpiOcular™ Eye Irritation Test (EIT)
according to OECD TG 492 -
protocol



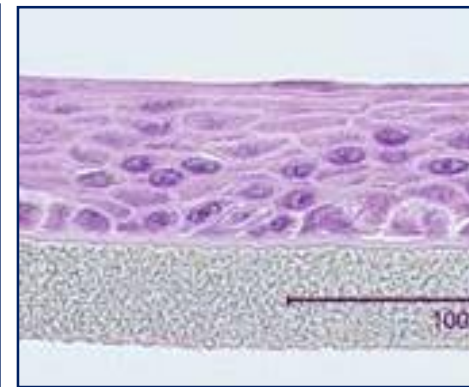
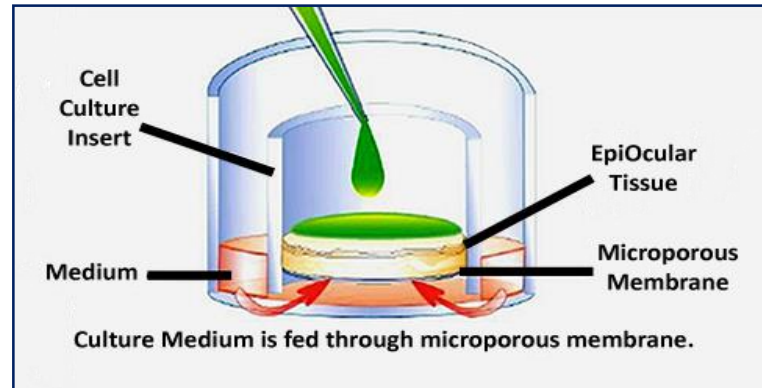
EpiOcular™ EIT kit (OCL-200-EIT kit)

Tissues are supplied as kit containing:

- 24 tissue inserts on transport agar,
- ASY culture medium (OCL-200-ASY),
- PC (methyl acetate)
- 6 well, 12 well and 24 well plates

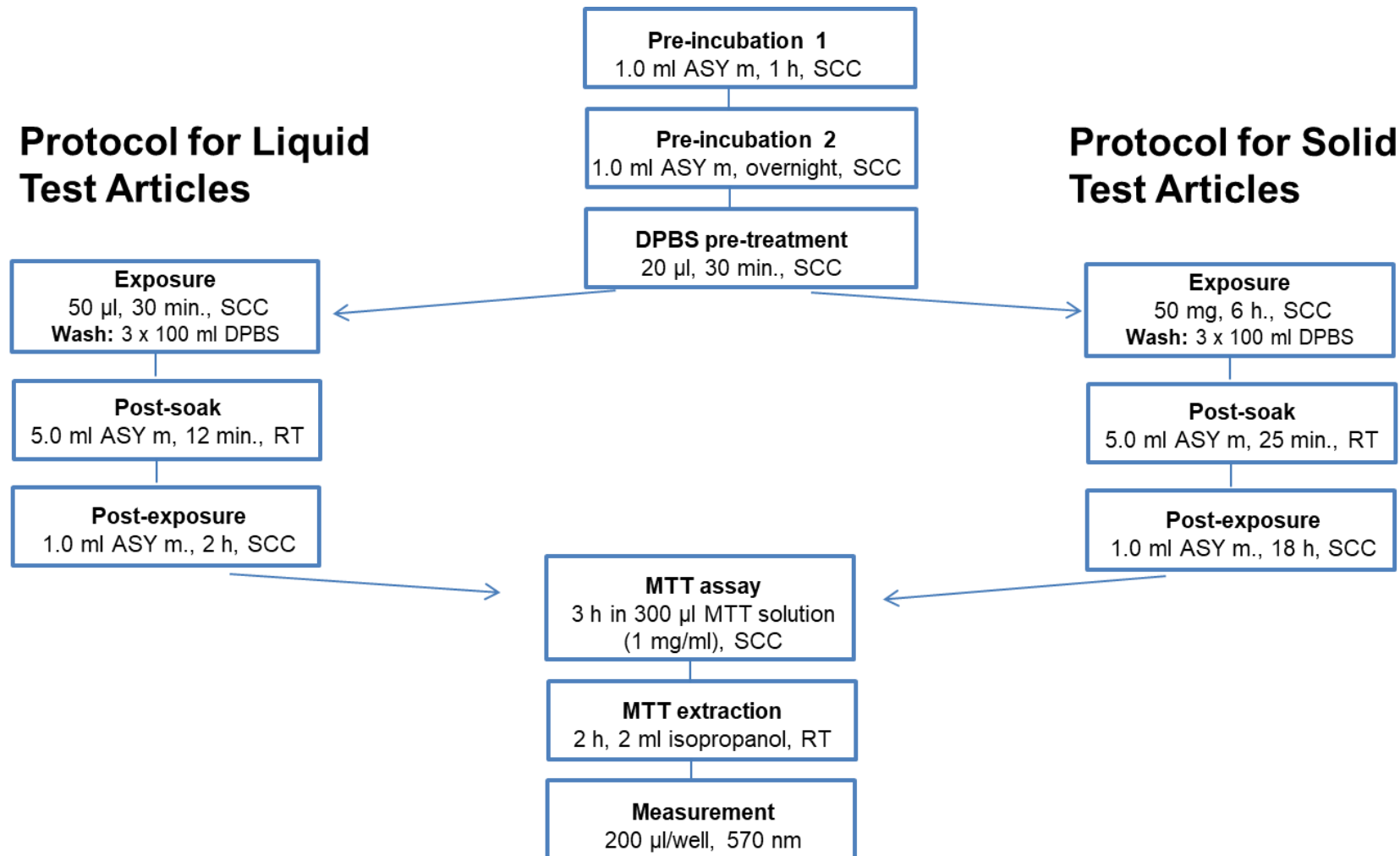
MTT - assay kit (MTT-100 kit):

- MTT concentrate (5 mg/ml) - 2ml
- MTT diluent (culture medium) - 8 ml
- MTT extractant (isopropanol) - 60 ml



Histology cross-section

Experimental design



Experimental design

Tissue 1	NC	PC	TA1	TA2	TA3	TA4
Tissue 2	NC	PC	TA1	TA2	TA3	TA4
Tissue 1	TA5	TA6	TA7	TA8	TA9	TA10
Tissue 2	TA5	TA6	TA7	TA8	TA9	TA10

The test is performed on a total of 2 tissues per test material, 2 tissues for negative control (NC), and 2 tissues for positive control (PC).

24 tissues = 10 Test chemicals (TC) + Negative (NC) and Positive control (PC)

Tissue 1	NC	PC	TA1	TA2	TA3	TA4	liquids
Tissue 2	NC	PC	TA1	TA2	TA3	TA4	
Tissue 1	NC	PC	TA5	TA6	TA7	TA8	solids
Tissue 2	NC	PC	TA5	TA6	TA7	TA8	

The test is performed on a total of 2 tissues per test material, 2 tissues for negative control, and 2 tissues for positive control.

12 tissues = 4 Test articles (TA) - liquids + Negative control (NC) and Positive control (PC)

12 tissues = 4 Test articles (TA) - solids + Negative control (NC) and Positive control (PC)

1. Pre-incubation

1/2

Before opening the OCL-200 kit, prefill all wells of four 6-well plates with 1.0 mL ASY (OCL-200-ASY) medium. Use the following plate design:

Negative control	Positive control	Test article 1
Negative control	Positive control	Test article 1

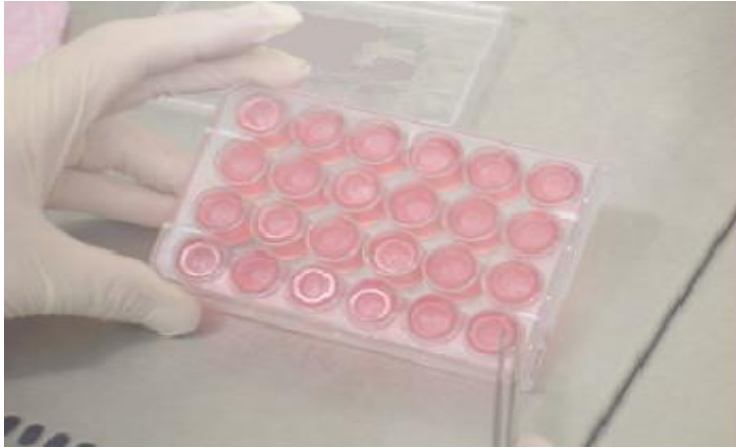
Test article 5	Test article 6	Test article 7
Test article 5	Test article 6	Test article 7

Test article 2	Test article 3	Test article 4
Test article 2	Test article 3	Test article 4

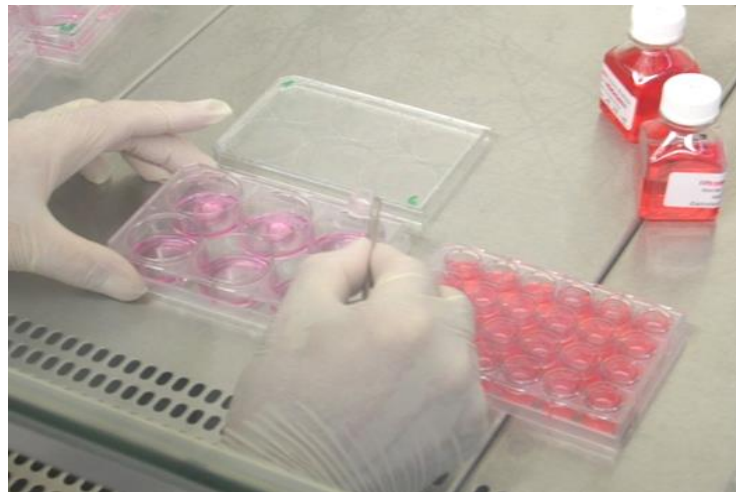
Test article 8	Test article 9	Test article 10
Test article 8	Test article 9	Test article 10

1. Pre-incubation

2/2



On day of receipt of the EpiOcular™ kit (usually Tuesday), tissues are transferred into 6-well plates pre-filled with ASY medium (OCL-200-ASY) (1.0 ml/well) and are conditioned 1 hour in the incubator (37°C, 5 % CO₂, humidified atmosphere).



After 1h, the ASY medium (OCL-200-ASY) is exchanged for the fresh one to fresh ASY medium (1.0 ml/well) and are further incubated overnight in the incubator (37°C, 5 % CO₂, humidified atmosphere).

2. DPBS pre-treatment

After the overnight incubation, the tissues are pre-wetted with 20 µl of DPBS and are incubated at standard culture conditions for **30 ± 2 min**.

3. Application of liquid test articles



The test is performed on a total of 2 tissues per test material, 2 tissue for negative control, and 2 tissues for positive control.

Negative control: sterile ultrapure H₂O

Positive control: methyl acetate

DOSE

liquids - 50 µl (undiluted test material)

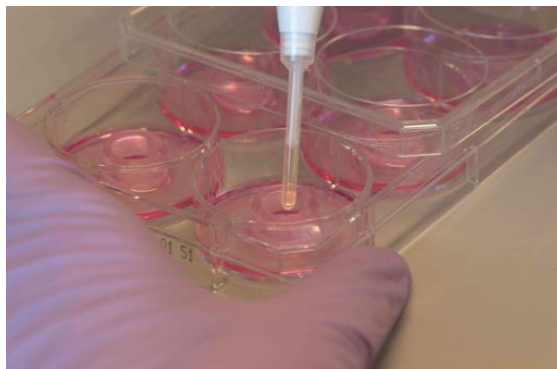
APPLICATION

liquids: pipetting

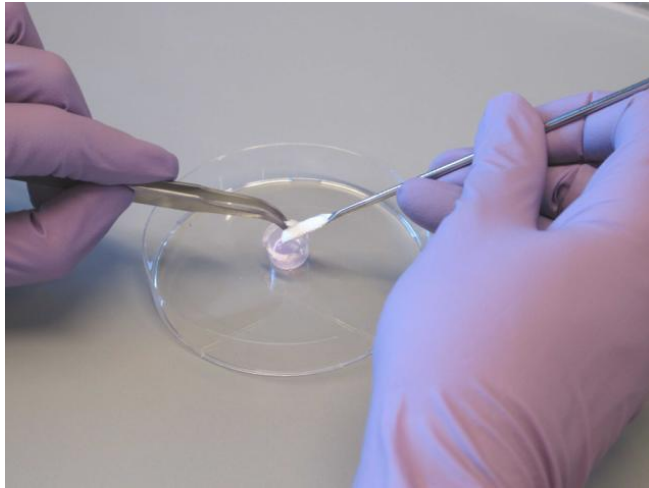
viscous liquids: positive displacement pipette

EXPOSURE TIME

30 ± 2 min



4. Application of solid test articles



The test is performed on a total of 2 tissues per test material, 2 tissue for negative control, and 2 tissues for positive control.

Negative control: sterile ultrapure H₂O

Positive control: methyl acetate

DOSE

NC and PC - 50 μ l

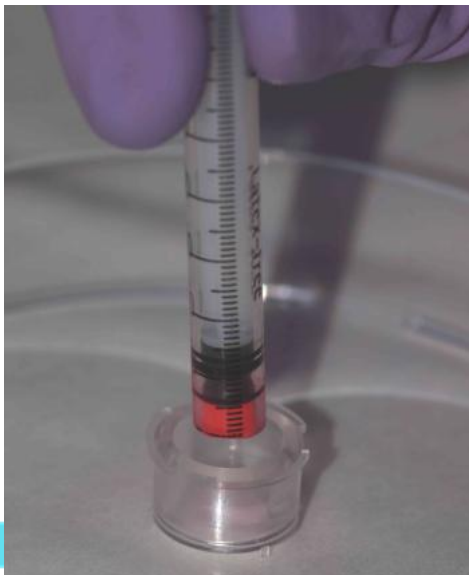
solids - 50 mg

APPLICATION

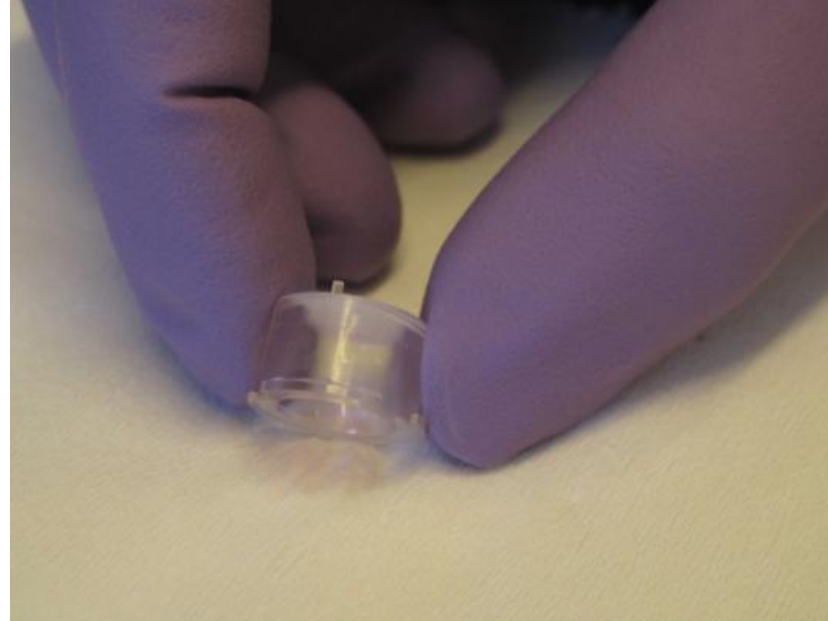
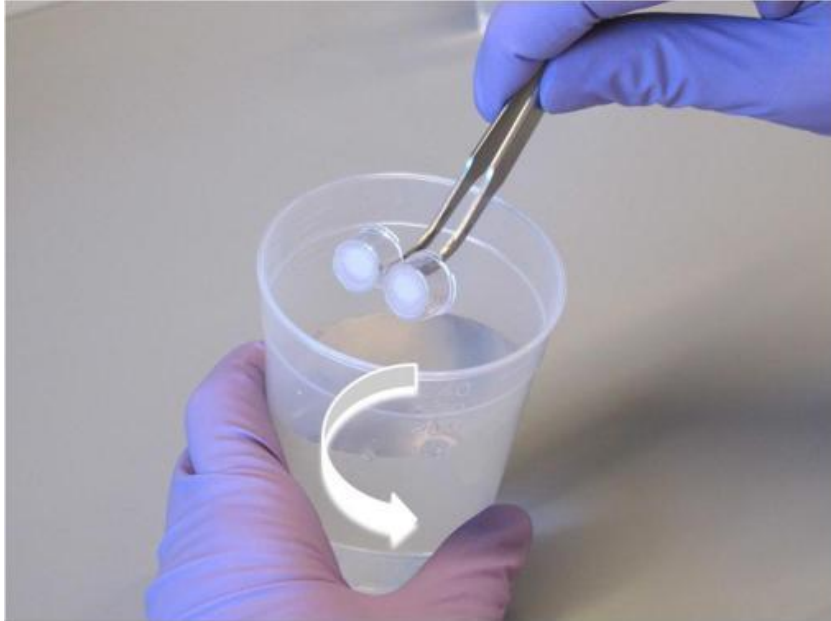
solids: spoon or syringe application

EXPOSURE TIME

6 h \pm 15 min



5. Washing procedure



After **30 min \pm 2 min** exposure to the **liquid** test articles or after **6 h \pm 15 min** exposure to the **solid** test articles, the tissues are washed 3 times with phosphate buffered saline (DPBS) to remove residual test material.

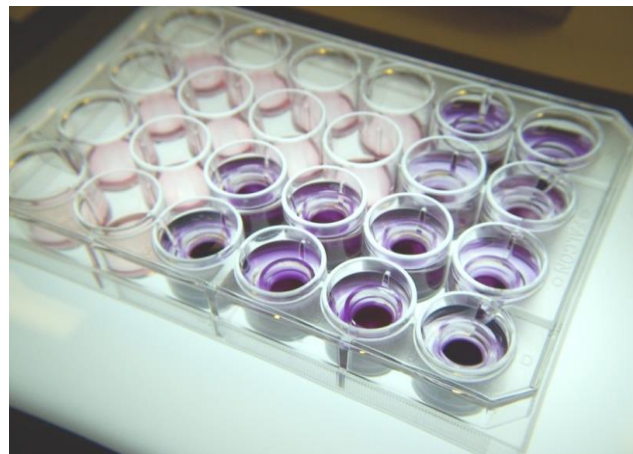
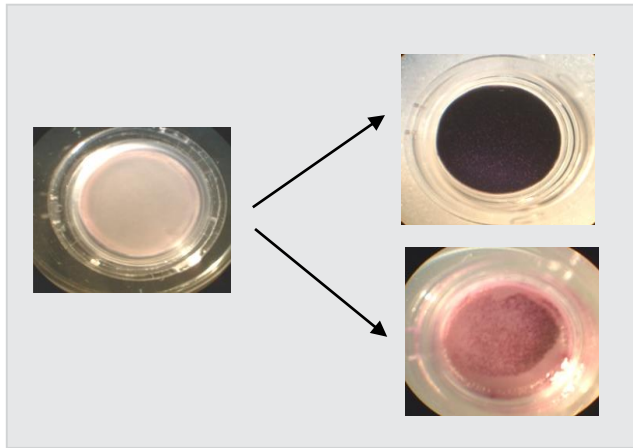
6. Post-soak and post-exposure

Rinsed and blotted inserts are transferred and immersed in 5 ml of ASY medium in a pre-labeled 12-well plates prefilled with 5 mL of ASY medium for post-soak for **12 min \pm 2 min** for **liquids** and **25 min \pm 2 min** for **solids**.

Afterwards, the medium is decanted off the tissue, the insert is blotted on the absorbent material, and transferred to the appropriate well of the pre-labelled 6-well plate containing 1 ml of ASY medium. The tissues are incubated for **120 min \pm 15 min** for **liquids** and **18 h \pm 0.25 h** for **solids** at standard culture conditions.



7. MTT assay and isopropanol extraction



After termination of post-exposure (**120 min \pm 15 min for liquids** and **18 h \pm 0.25 h for solids**), tissues are transferred into 24 well plates prefilled with 0.3 mL MTT medium. Tissue are incubated with MTT for **3 h \pm 10 min** (37°C, 5% CO₂, humidified atmosphere) protected from light.

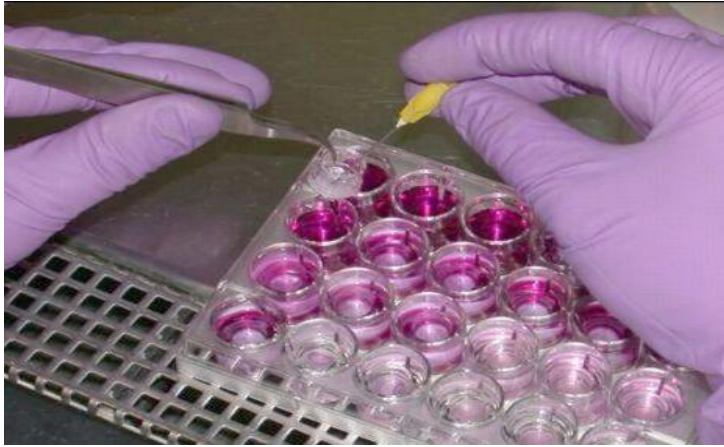
Viable cells/tissues will convert yellow MTT into purple/blue formazan product. Non-viable skin models will remain unstained.

For liquids: After incubation with MTT is completed, tissues are transferred into new 24 -well plate and formazan is extracted with 2 mL isopropanol (2 h at room temperature or overnight at 4°C).

For solids: After incubation with MTT is completed, tissues are transferred into 6-well plate containing 2 ml of isopropanol so that no isopropanol is floating in the insert. (2 h at room temperature or overnight at 4°C).

Plated should be sealed (e.g. with parafilm) to avoid evaporation of isopropanol.

8. Preparation of 96 well plate for OD measurement



After the extraction period is completed, inserts are pierced with an injection needle in the case of **liquids**.

For **solids**, the tissues should not be pierced.

Extract will run into the well from which the insert was taken. Afterwards, the insert can be discarded. Plates are placed on a shaker for 15 minutes until solution is homogeneous.



Per each tissue $2 \times 200\mu\text{L}$ aliquots of the blue formazan solution are transferred into a 96-well flat bottom microtiter plate

For the measurement in a 96 well plate, use exactly the plate design given in the spreadsheet for calculation.

9. Measurement

Fixed plate design

	1	2	3	4	5	6	7	8	9	10	11	12	
A	NC	PC	C1	C2	C3	C4	C5	C6	C7	C8	C9	C10	Tissue1
B	NC	PC	C1	C2	C3	C4	C5	C6	C7	C8	C9	C10	
C	NC	PC	C1	C2	C3	C4	C5	C6	C7	C8	C9	C10	Tissue2
D	NC	PC	C1	C2	C3	C4	C5	C6	C7	C8	C9	C10	
E	C11	C12	C13	C14	C15	C16	C17	C18	C19	C20	BLANK	BLANK	Tissue1
F	C11	C12	C13	C14	C15	C16	C17	C18	C19	C20	BLANK	BLANK	
G	C11	C12	C13	C14	C15	C16	C17	C18	C19	C20	BLANK	BLANK	Tissue2
H	C11	C12	C13	C14	C15	C16	C17	C18	C19	C20	BLANK	BLANK	

Read optical density (OD) in a plate spectrophotometer at **570 nm, without reference filter.**

The OD of the formazan can be read at in a range of **550 nm – 590nm.**



Acceptability ranges for NC OD values in EpiOcular EIT are **> 0.8 and < 2.8.**

EpiOcular Eye Irritation Test Spreadsheet - import

EpiOcular - Eye Irritation Test (OCL-200-EIT)

Correspondent informations of MD's	
Exp. no.:	V16-5263
Tissue-lot no.:	23499 K1 F
Date:	27-Oct-16
Operator:	SN LV

Negative control	NC (DI H2O)		
Positive control	PC (methyl acetate)		
Test Chemical No. 1	1234	Test Chemical No. 11	7777
Test Chemical No. 2	2345	Test Chemical No. 12	8888
Test Chemical No. 3	1234	Test Chemical No. 13	9999
Test Chemical No. 4	2345	Test Chemical No. 14	8877
Test Chemical No. 5	1234	Test Chemical No. 15	7777
Test Chemical No. 6	2345	Test Chemical No. 16	8888
Test Chemical No. 7	1234	Test Chemical No. 17	9999
Test Chemical No. 8	2345	Test Chemical No. 18	8877
Test Chemical No. 9	2345	Test Chemical No. 19	6666
Test Chemical No. 10	2345	Test Chemical No. 20	3232

FIXED DESIGN OF 96 WELL PLATE

PLATE 1

	1	2	3	4	5	6	7	8	9	10	11	12	
A	NC	PC	1234	2345	1234	2345	1234	2345	1234	2345	2345	2356	Tissue1
B	NC	PC	1234	2345	1234	2345	1234	2345	1234	2345	2345	2356	
C	NC	PC	1234	2345	1234	2345	1234	2345	1234	2345	2345	2356	Tissue2
D	NC	PC	1234	2345	1234	2345	1234	2345	1234	2345	2345	2356	
E	7777	8888	9999	8877	7777	8888	9999	8877	6666	3232	BLANK	BLANK	Tissue1
F	7777	8888	9999	8877	7777	8888	9999	8877	6666	3232	BLANK	BLANK	
G	7777	8888	9999	8877	7777	8888	9999	8877	6666	3232	BLANK	BLANK	Tissue2
H	7777	8888	9999	8877	7777	8888	9999	8877	6666	3232	BLANK	BLANK	

IMPORT:

PLATE 1

	1	2	3	4	5	6	7	8	9	10	11	12	
A	2.142	0.840	1.906	1.853	1.490	1.787	1.812	0.510	1.578	1.523	1.150	1.457	Tissue1
B	2.147	0.820	1.912	1.854	1.536	1.776	1.817	0.490	1.582	1.524	1.208	1.446	
C	2.282	0.848	1.876	1.757	1.932	2.018	1.952	0.318	1.546	1.427	1.802	1.688	Tissue2
D	2.243	0.658	1.821	1.730	1.851	2.039	1.913	0.328	1.491	1.400	1.521	1.709	
E	0.109	0.119	0.101	1.754	0.198	0.309	0.319	0.301	1.954	0.511	0.037	0.038	Tissue1
F	0.109	0.114	0.104	1.739	0.209	0.309	0.314	0.304	1.939	0.512	0.038	0.037	
G	0.223	0.333	0.232	1.886	0.223	0.423	0.411	0.411	1.886	0.423	0.038	0.038	Tissue2
H	0.224	0.334	0.233	1.566	0.288	0.424	0.423	0.423	1.786	0.278	0.038	0.037	

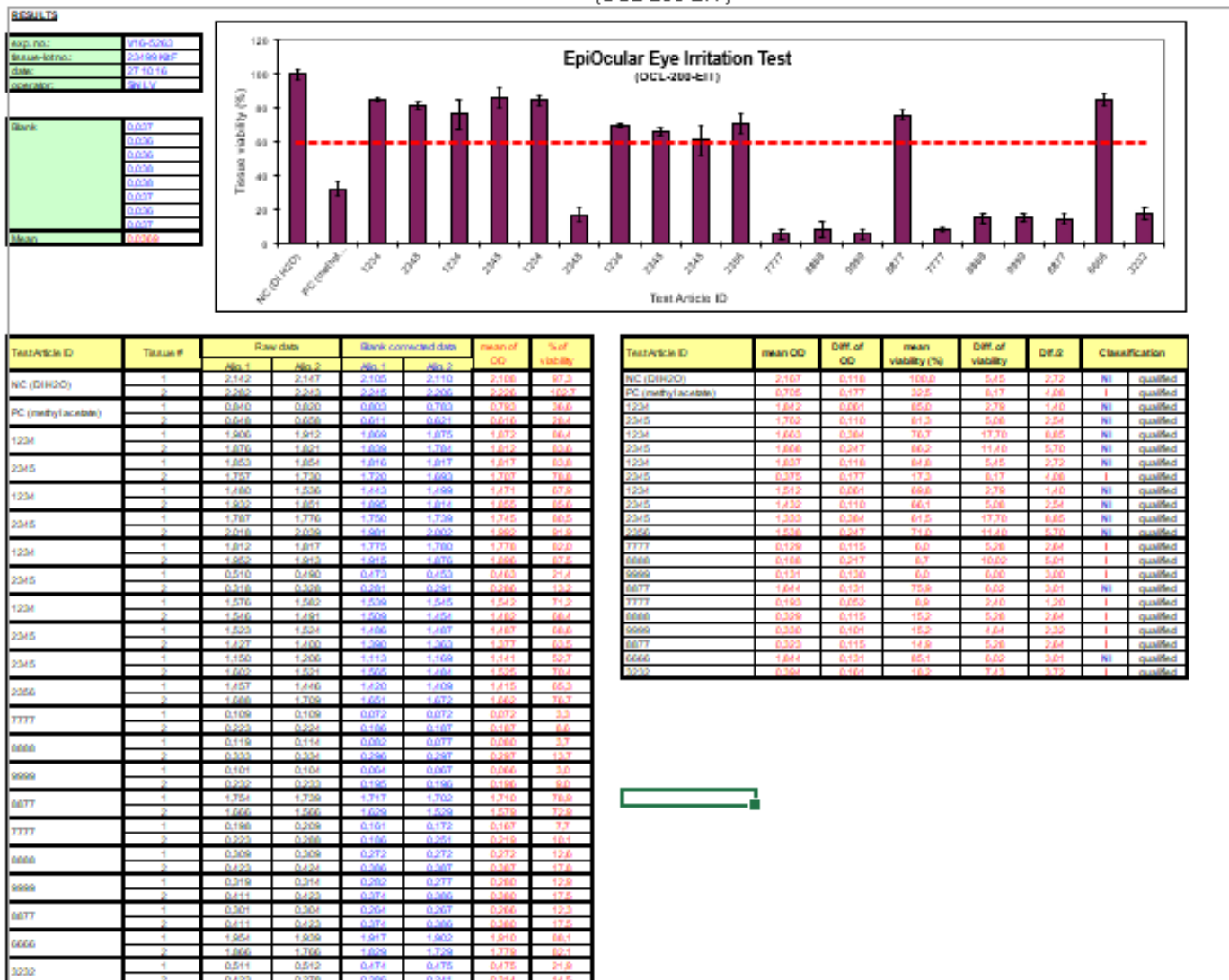
REMARKS

Date:

Performed by:

EpiOcular Eye Irritation Test Spreadsheet - calculation

EpiOcular Eye Irritation Test
(OCL-200-ET)



JoVE video-protocol EpiOcular EIT (free access)

<http://www.jove.com/video/52979/eye-irritation-test-eit-for-hazard-identification-eye-irritating>

B N F M BE ENG C Bh E DB SE

BE Eye Irritation Test (EIT) for Hazard Identification of Eye Irritating Chemicals using Reconstructed Human Cornea-like Epithelial (RhCE) Tissue Model

Yulia Kaluzhny¹, Helena Kandárová², Laurence d'Argembeau-Thornton¹, Paul Kearney¹, Mitchell Klausner¹

¹MatTek Corporation, ²MatTek In Vitro Life Science Laboratories

Article

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Metrics

2



0:05 Title

1:43 Reconstructed Human Cornea-like Epithelial (RhCE) Tissue Treatment Preparation (Day 0)

3:08 Pre-treatment and Test Article Exposure (Day 1)

4:42 Rinsing

6:13 MTT Viability Assay (Liquid Test Day 1; Solid Test Day 2)

8:19 Results: Representative EIT Results for 10 Test Articles

9:36 Conclusion

This article is **Open Access**.



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Translate text to:

Summary

We have developed an eye irritation test which utilizes a three dimensional reconstructed human cornea-like epithelial (RhCE) tissue model. The test is able to discriminate between ocular irritant and corrosive materials (GHS Categories 1 and 2 combined) and those that do not require labeling (GHS No Category).

Date Published: 8/23/2015, **Issue 102**; doi: [10.3791/52979](https://doi.org/10.3791/52979)

Keywords: Bioengineering, Issue 102, alternative methods, cornea, EpiOcular, EURL ECVAM, eye irritation, in vitro, MTT viability assay, ocular irritation, REACH, reconstructed ocular tissue model, RhCE



Thank you very much for your attention!

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