

EpiOcular[™] EIT kit (OCL-200-EIT kit)

Tissues are supplied as kit containing:

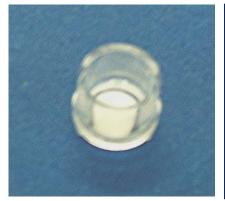
24 tissue inserts on transport agar, ASY culture medium (OCL-200-ASY), PC (methyl acetate) 6 well, 12 well and 24 well plates

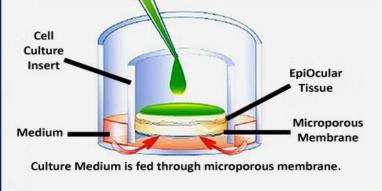
MTT - assay kit (MTT-100 kit):

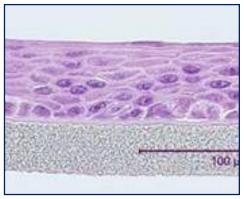
MTT concentrate (5 mg/ml) - 2ml MTT diluent (culture medium) - 8 ml MTT extractant (isopropanol) - 60 ml

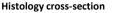






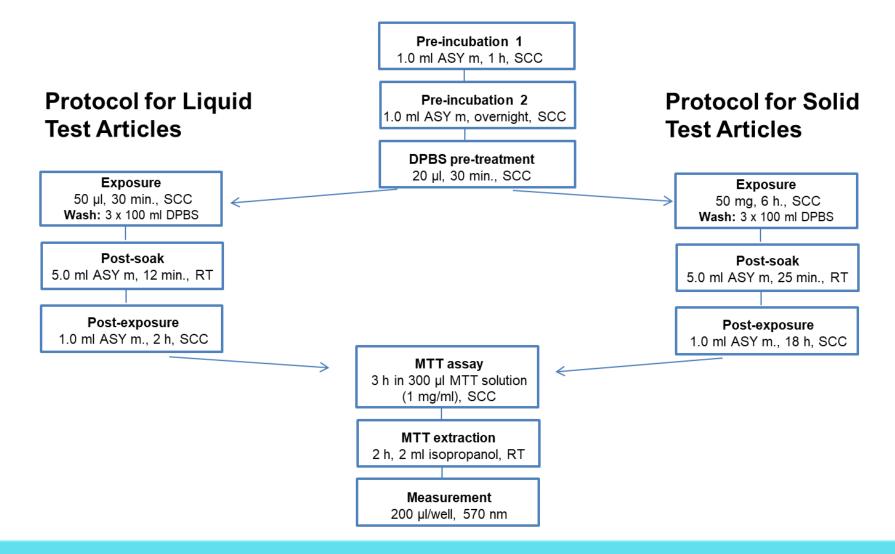






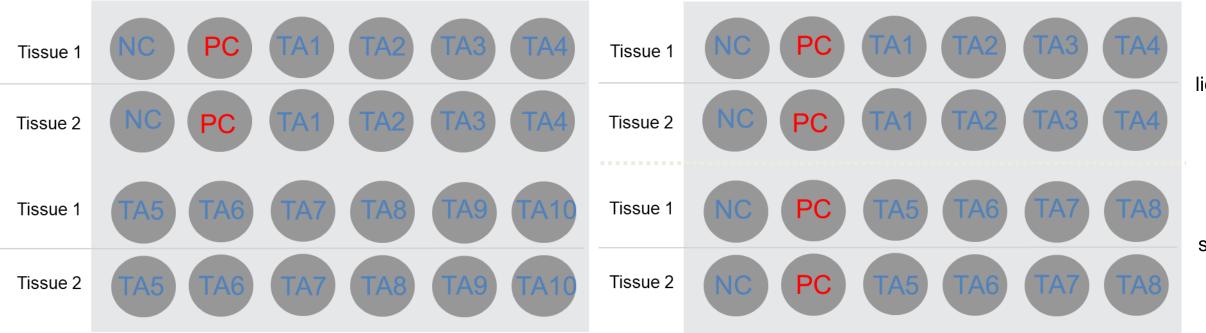


Experimental design





Experimental design



The test is performed on a total of 2 tissues per test material, 2 tissues for negative control (NC), and 2 tissues for positive control (PC).

24 tissues = 10 Test chemicals (TC) + Negative (NC) and Positive control (PC)

The test is performed on a total of 2 tissues per test material, 2 tissues for negative control, and 2 tissues for positive control.

12 tissues = 4 Test articles (TA) - liquids + Negative control (NC) and Positive control (PC)

12 tissues = 4 Test articles (TA) - solids + Negative control (NC) and Positive control (PC)



liquids

solids

Before opening the OCL-200 kit, prefill all wells of four 6-well plates with 1.0 mL ASY (OCL-200-ASY) medium. Use the following plate design:

Negative control	Positive control	Test article 1
Negative control	Positive control	Test article 1

Test article 5	Test article 6	Test article 7
Test article 5	Test article 6	Test article 7

Test article 2	Test article 3	Test article 4
Test article 2	Test article 3	Test article 4

Test article 8	Test article 9	Test article 10
Test article 8	Test article 9	Test article 10



1. Pre-incubation



On day of receipt of the EpiOcular[™] kit (usually Tuesday), tissues are transferred into 6-well plates pre-filled with ASY medium (OCL-200-ASY) (1.0 ml/well) and are conditioned 1 hour in the incubator (37°C, 5 % CO₂, humidified atmosphere).



After 1h, the ASY medium (OCL-200-ASY) is exchanged for the fresh one to fresh ASY medium (1.0 ml/well) and are further incubated overnight in the incubator (37°C, 5 % CO_2 , humidified atmosphere).



2. DPBS pre-treatment

After the overnight incubation, the tissues are pre-wetted with 20 μ l of DPBS and are incubated at standard culture conditions for **30 ± 2 min.**

3. Application of liquid test articles



The test is performed on a total of 2 tissues per test material, 2 tissue for negative control, and 2 tissues for positive control.

Negative control: sterile ultrapure H2O

Positive control: methyl acetate



liquids - 50 µl (undiluted test material)

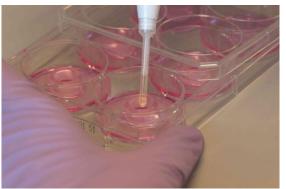


liquids: pipetting

viscous liquids: positive displacement pipette

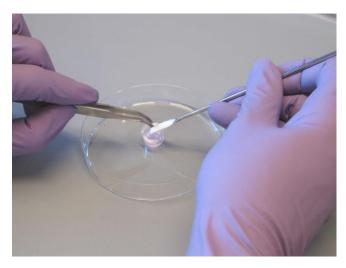
EXPOSURE TIME

30 ± 2 min





4. Application of solid test articles



The test is performed on a total of 2 tissues per test material, 2 tissue for negative control, and 2 tissues for positive control.

Negative control: sterile ultrapure H2O

Positive control: methyl acetate

DOSE

NC and PC - 50 µl solids - 50 mg

APPLICATION

solids: spoon or syringe application

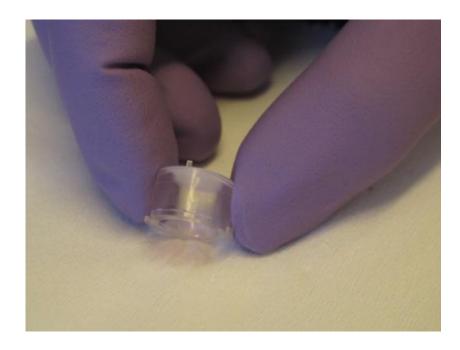
EXPOSURE TIME 6 h ± 15 min





5. Washing procedure





After 30 min \pm 2 min exposure to the liquid test articles or after 6 h \pm 15 min exposure to the **solid** test articles, the tissues are washed 3 times with phosphate buffered saline (DPBS) to remove residual test material.



6. Post-soak and post-exposure

Rinsed and blotted inserts are transferred and immersed in 5 ml of ASY medium in a pre-labeled 12-well plates prefilled with 5 mL of ASY medium for post-soak for 12 min ± 2 min for liquids and 25 min ± 2 min for solids.

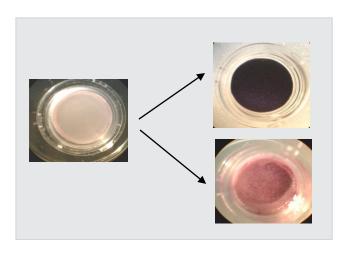
Afterwards, the medium is decanted off the tissue, the insert is blotted on the absorbent material, and transferred to the appropriate well of the pre-labelled 6-well plate containing 1 ml of ASY medium. The tissues are incubated for **120 min ± 15 min** for **liquids** and **18 h ± 0.25 h** for **solids** at standard culture conditions.







7. MTT assay and isopropanol extraction





After termination of post-exposure (120 min \pm 15 min for liquids and 18 h \pm 0.25 h for solids), tissues are transferred into 24 well plates prefilled with 0.3 mL MTT medium. Tissue are incubated with MTT for 3 h \pm 10 min (37°C, 5% CO₂, humidified atmosphere) protected from light.

Viable cells/tissues will convert yellow MTT into purple/blue formazan product. Non-viable skin models will remain unstained.

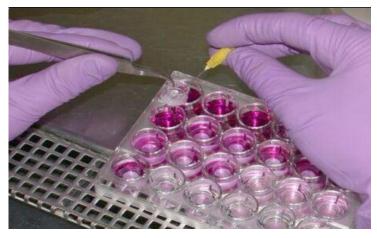
For liquids: After incubation with MTT is completed, tissues are transferred into new 24 -well plate and formazan is extracted with 2 mL isopropanol (2 h at room temperature or overnight at 4°C).

For solids: After incubation with MTT is completed, tissues are transferred into 6-well plate containing 2 ml of isopropanol so that no isopropanol is floating in the insert. (2 h at room temperature or overnight at 4°C).

Plated should be sealed (e.g. with parafilm) to avoid evaporation of isopropanol.



8. Preparation of 96 well plate for OD measurement





After the extraction period is completed, inserts are pierced with an injection needle in the case of **liquids**.

For **solids**, the tissues should not be pierced.

Extract will run into the well from which the insert was taken. Afterwards, the insert can be discarded. Plates are placed on a shaker for 15 minutes until solution is homogeneous.

Per each tissue $2 \times 200 \mu L$ aliquots of the blue formazan solution are transferred into a 96-well flat bottom microtiter plate

For the measurement in a 96 well plate, <u>use exactly the plate</u> <u>design given in the spreadsheet for calculation</u>.



9. Measurement

Fixed plate design

	12	11	10	9	8	7	6	5	4	3	2	1	
T:4	C10	C9	C8	C7	C6	C5	C4	C3	C2	C1	PC	NC	А
Tissue1	C10	C9	C8	C7	C6	C5	C4	C3	C2	C1	PC	NC	В
T0	C10	C9	C8	C7	C6	C5	C4	C3	C2	C1	PC	NC	С
Tissue2	C10	C9	C8	C7	C6	C5	C4	C3	C2	C1	PC	NC	D
T1	BLANK	BLANK	C20	C19	C18	C17	C16	C15	C14	C13	C12	C11	E
Tissue1	BLANK	BLANK	C20	C19	C18	C17	C16	C15	C14	C13	C12	C11	F
T0	BLANK	BLANK	C20	C19	C18	C17	C16	C15	C14	C13	C12	C11	G
Tissue2	BLANK	BLANK	C20	C19	C18	C17	C16	C15	C14	C13	C12	C11	Н

Read optical density (OD) in a plate spectrophotometer at **570 nm**, **without reference filter**.

The OD of the formazan can be read at in a range of **550 nm – 590nm**.



Acceptability ranges for NC OD values in EpiOcular EIT are > 0.8 and < 2.8.



EpiOcular Eye Irritation Test Spreadsheet - import

EpiOcular - Eye Irritation Test (OCL-200-EIT)

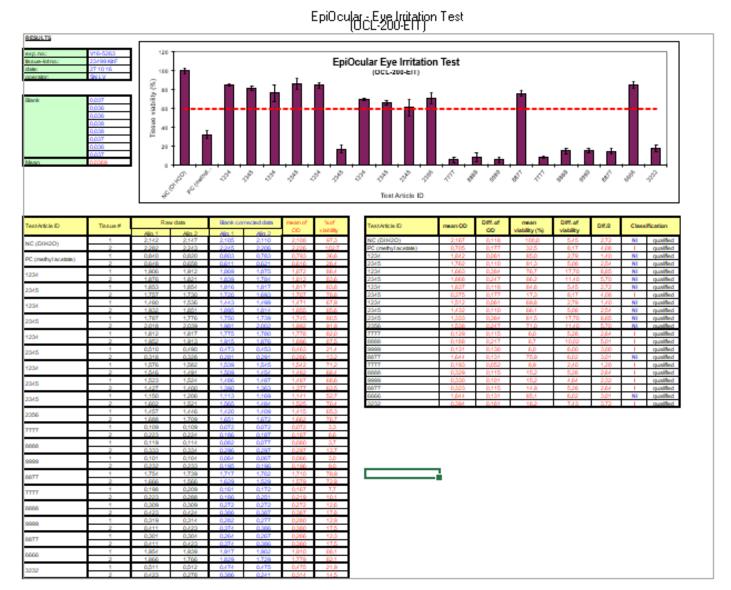
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Date: Performed by:

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EpiOcular Eye Irritation Test Spreadsheet - calculation



JoVE video-protocol EpiOcular EIT (free access)

http://www.jove.com/video/52979/eye-irritation-test-eit-for-hazard-identification-eye-irritating





