



In vitro experimental system  
relevance and factors influencing  
the outcome of the study.

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# Cosmetic ingredient evaluation: Setting the Scene

Development of New Approach Methodologies (**NAMs**) by using human cells, with monolayer (co) cultures to more complex cell models (3D or organ-on-chip), together with computational methods (e.g. QIVIVE, PBK),

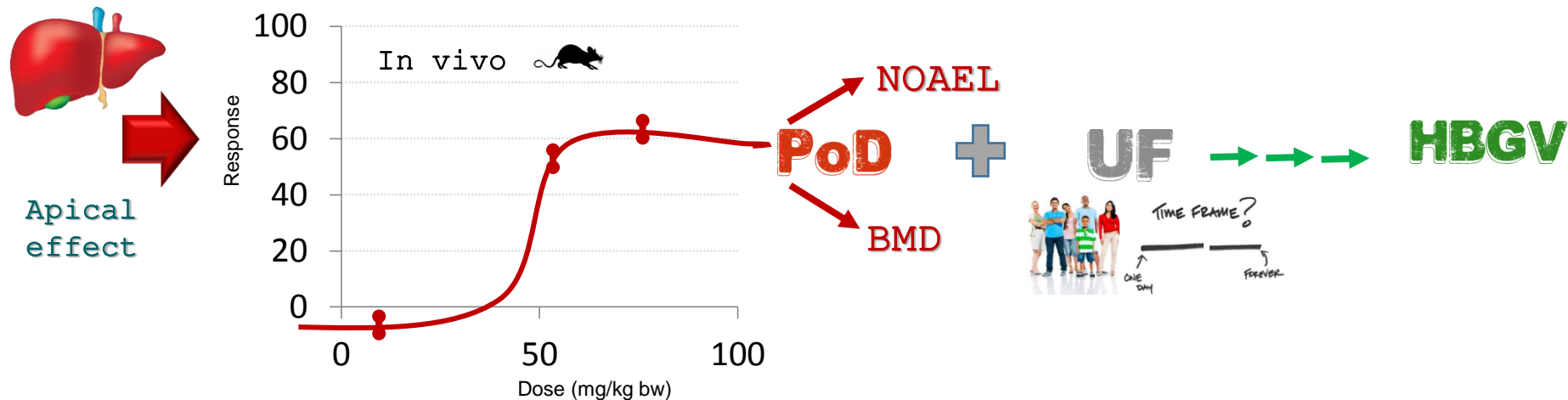


reduction of the number of animals used in different research and regulatory sectors.

- Cosmetic ingredients evaluation by using NAM-based strategies is an essential step to ensure the safety profile of cosmetics (Scientific Committee on Consumer Safety-SCCS).
- A huge challenge to guarantee the absence of risk **ONLY** based on the current available methods.

# Next Generation Risk Assessment NGRA Development New Cosmetic Ingredient

## From Classical Toxicology

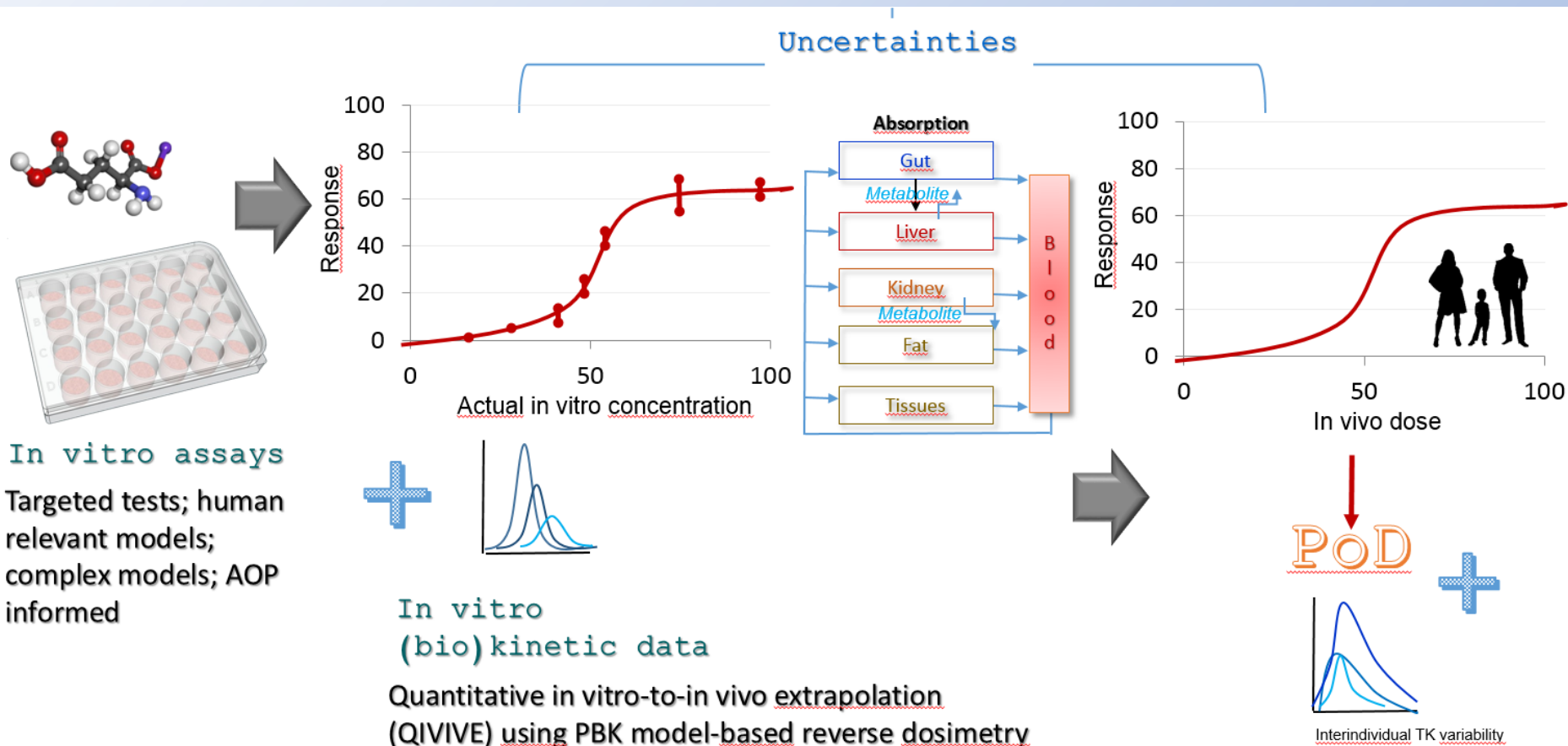


Modified from:  
Ingenbleek et al., 2021; DOI:10.1039/9781839160431-00001;  
Levorato et al., 2019 Current Opinion in Food Science

Di Consiglio, et al., Toxicol Lett., (2021);  
Vichi, et al., Toxicol Lett., (2021);  
Algharably, Di Consiglio, et al., Arch Toxicol., (2021);  
Di Consiglio, et al., Reprod Toxicol. (2020)

# Next Generation Risk Assessment **NGRA**

## Development New Cosmetic Ingredient



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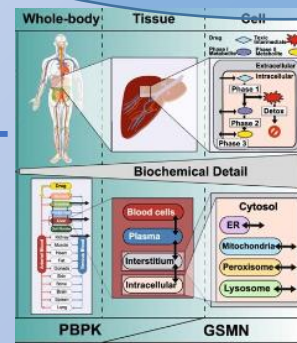
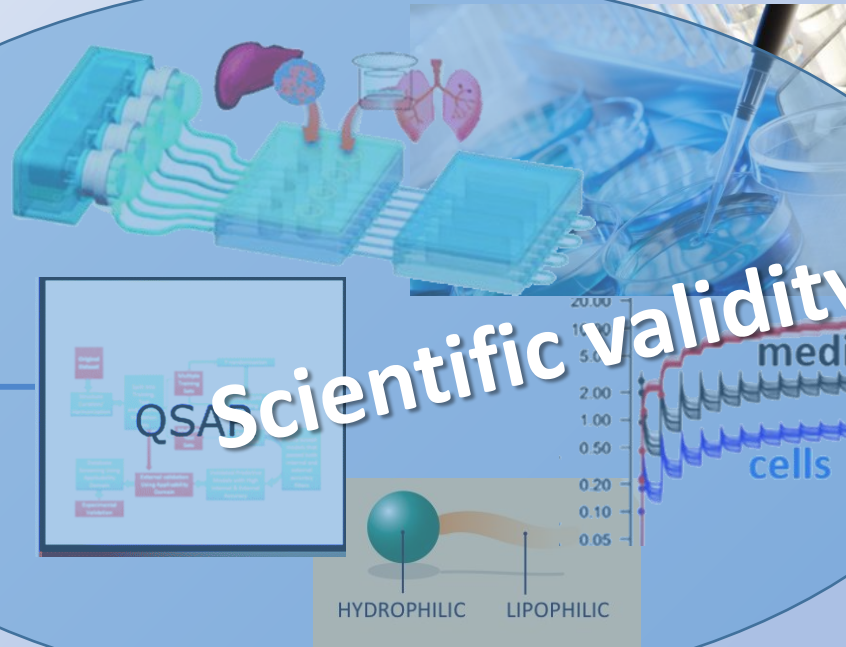
# Cosmetic ingredient evaluation: Next Generation Risk Assessment

MOS

Actual  
PoD

Exposure

Scientific validity



2021- SCCS/1628/21

Modified from Rogiers et al., *Toxicology* 436 (2020)

# Scientific validity of a test method

Evaluating the capacity of a test to protect human health:

how well it correlates with reliable information on activity of chemicals in humans.

**≠ piece of the evidence → ≠ NAMs**

- ✓ scientifically valid;
- ✓ properly developed and adequately described;
- ✓ “fit-for-purpose” performance;
- ✓ adequate scientific quality standards: standardization, relevance, specificity, sensibility

2018- *Guidance Document on Good In Vitro Method Practices (GIVIMP)*, OECD Series on Testing and Assessment, No. 286, <https://doi.org/10.1787/9789264304796-en>

2022- *Guidance document on Good Cell and Tissue Culture Practice 2.0 (GCCP 2.0)*, ALTEX, 39(1), pp. 30–70. <https://doi: 10.14573/altex.2111011>

2021 - *Guidance document on the characterisation, validation and reporting of PBK models for regulatory purposes*, OECD Series on Testing and Assessment, No. 331

# What are the goals?



**BY** Improving performing and *reporting*\*;

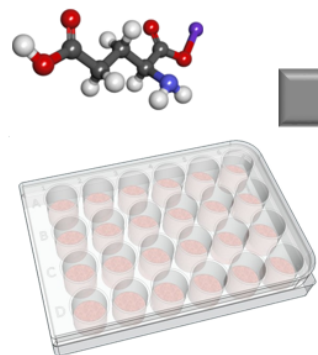
**BY** Improving implementation of requirements to consider all available information.

**Regulatory accepted** and **standardised** in vitro methods as internationally recognised OECD TG

**Academic data** used in regulatory assessments: helping researchers to design, perform and report studies, in order to facilitate regulatory acceptance

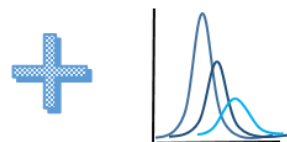
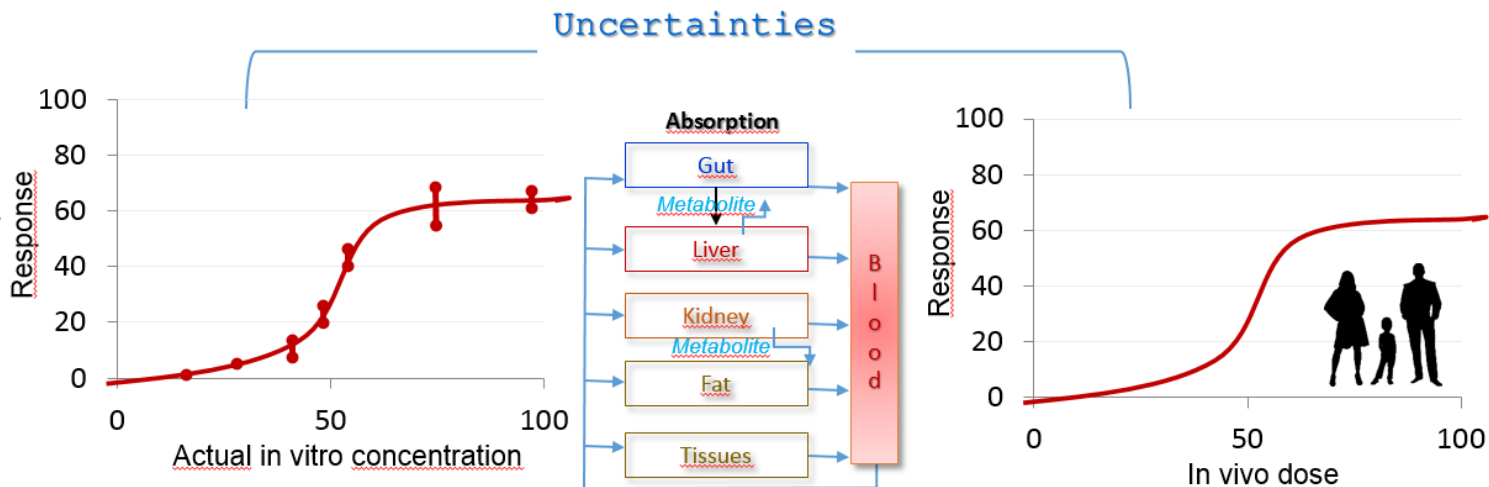
*\*2014, Guidance Document For Describing Non-guideline In Vitro Test Methods  
OECD Series on Testing and Assessment, No. 211*

# QIVIVE



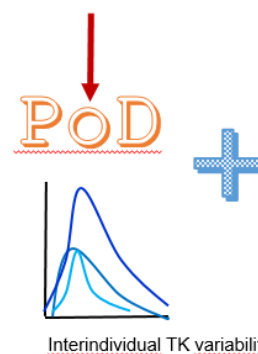
## In vitro assays

Targeted tests; human relevant models; complex models; AOP informed



## In vitro (bio)kinetic data

Quantitative in vitro-to-in vivo extrapolation (QIVIVE) using PBK model-based reverse dosimetry





Broadly defined as a **quantitative or qualitative transposition of in vitro experimental data** to predict in vivo effects, refer to :

1. The **estimation** of the **in vivo ADME**, based on parameters measured in vitro, often used for constructing PBPK models;
2. The process of converting an **in vitro concentration** associated with **bioactivity** to an external exposure level (i.e. **reverse dosimetry**) → use of PBPK model to determine an exposure level that leads to a tissue plasma concentration = to the in vitro concentration.

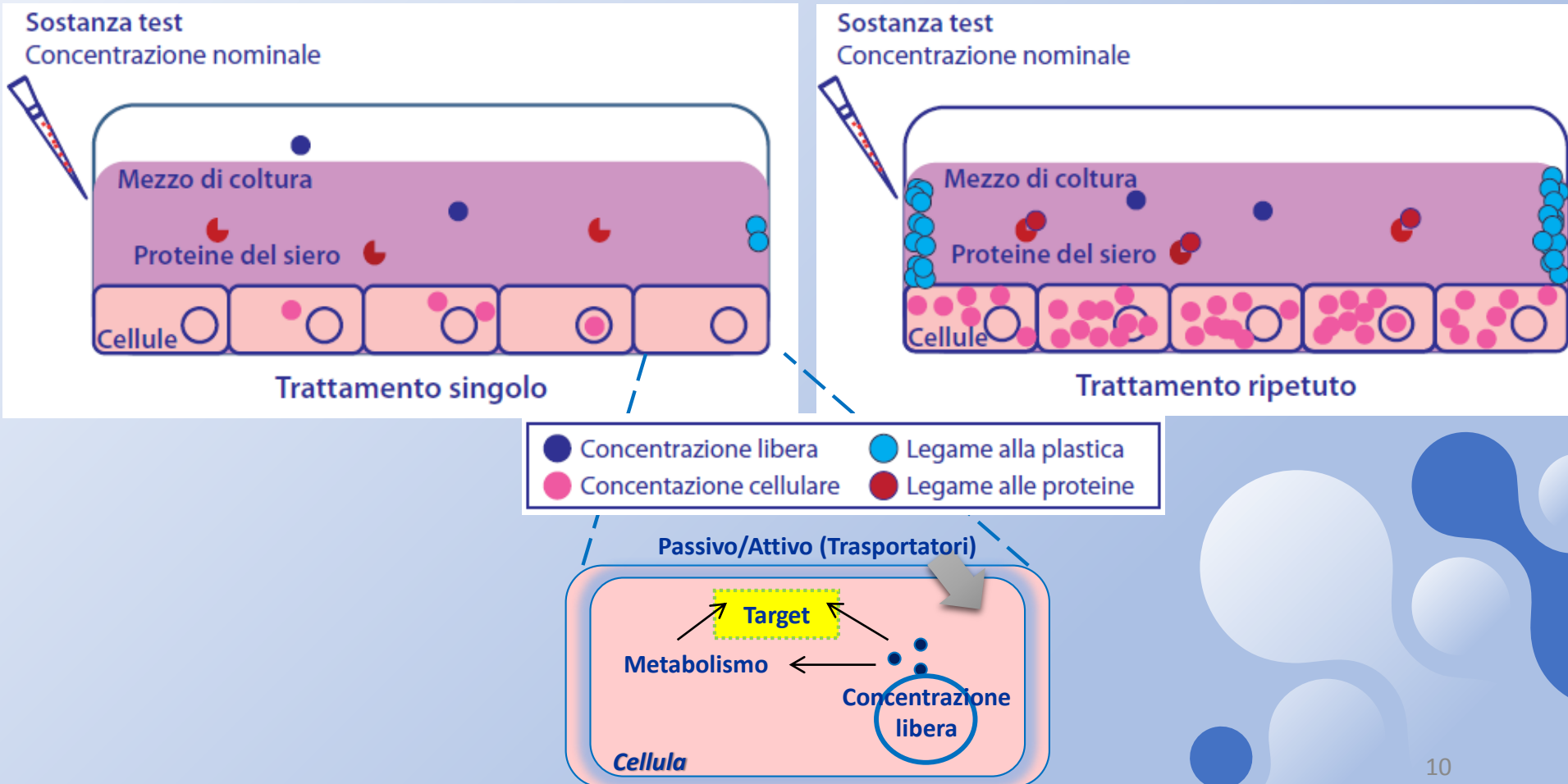
**The predicted exposure level  
can then be compared with the actual or estimated  
human exposures to estimate potential  
health risks**



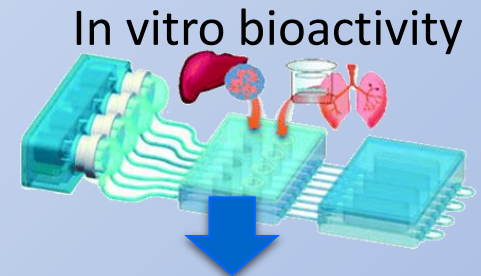
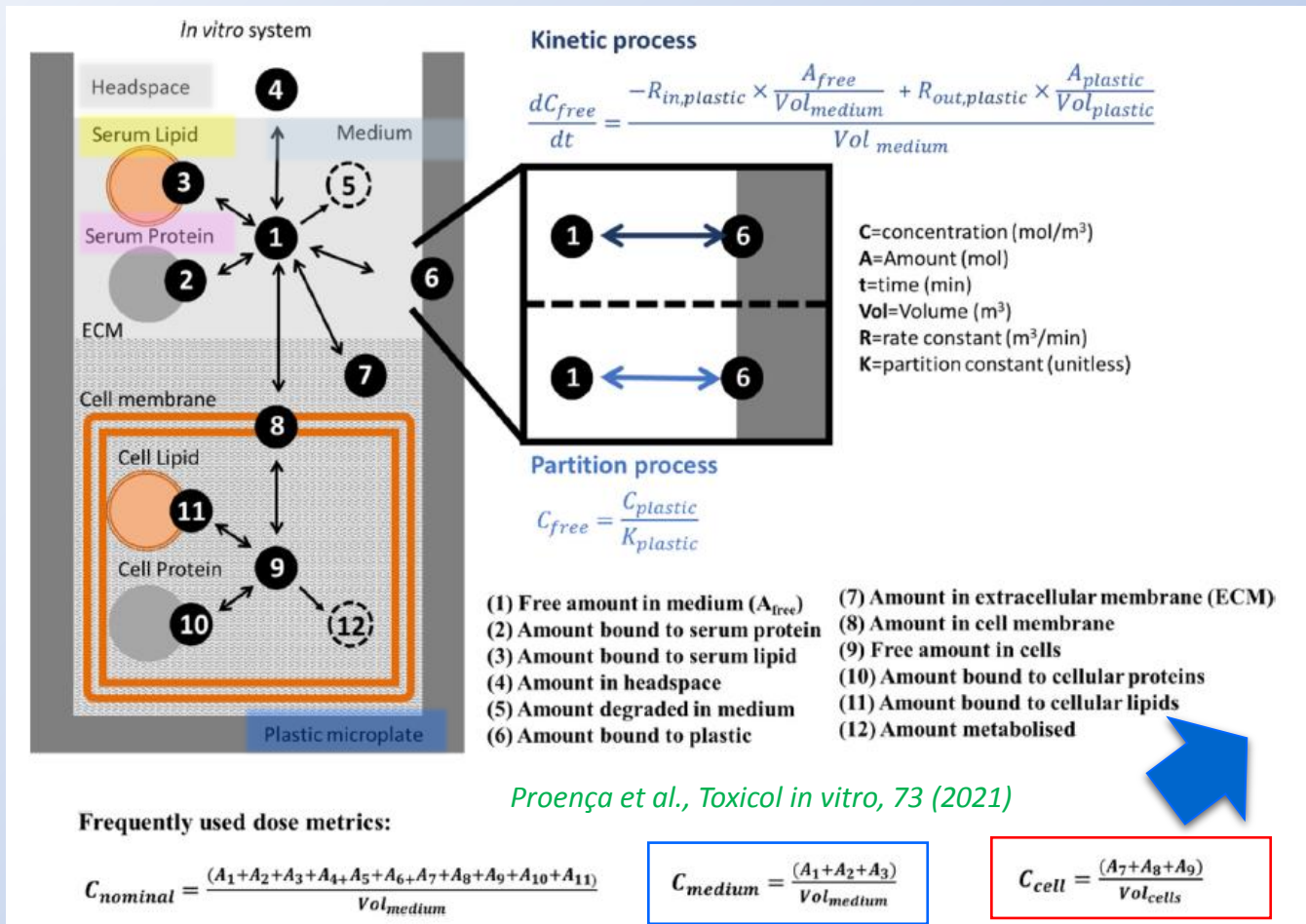
Don't  
Forget!!

One of the key barriers for the acceptance of in vitro toxicity testing data ? Failure to relate the nominal concentration to a relevant in vivo exposure level.

## In vitro system complexity: Biokinetics

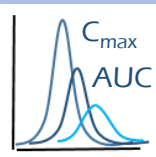


# Biokinetic factors influencing the outcome of the study

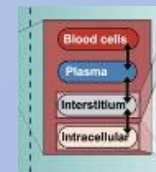


In vitro concentration  
+  
kinetic factors

In vivo  
**plasma** or  
**tissue**  
concentration



PBK



External dose



# In vitro biokinetics: Step-wise procedure for determination of chemical cellular bioavailability.

**In vitro kinetic factors to be considered:**

Chemical solubility in aqueous, medium and vehicle

Chemical stability in aqueous, medium and vehicle

Cross-contamination among wells

Non specific bindings

- with serum;
- with plastic devices
- with matrix used to culture the cells

**Bio Effective  
Concentration**

**Repeated  
dosing**

Time dependence

- Intracellular uptake
- Amount in the medium
- Biotransformation
- Bioaccumulation

**Metabolic  
capacity**  
**Transporter  
expression**

*Modified from Di Consiglio et al., Reproductive Toxicology 98 (2020)*

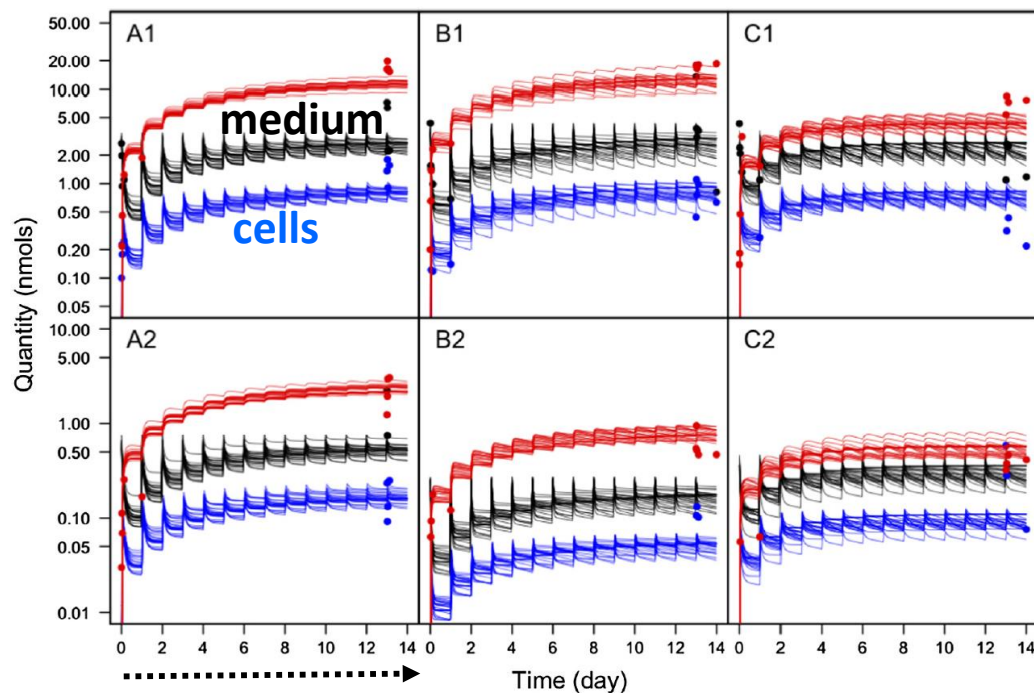


# Biokinetics “FEASIBLE” after Repeated dosing

**Lipophilic substance:**

(log  $P_{O/W}$  7.57); high protein binding (96–99%); extensive tissue distribution

**Repeated dosing** →



*Pomponio et al., Toxicology in Vitro 30 (2015) 36–51*

*Algharably, et al., Archives of Toxicology, 93, (2019) 615-621.*

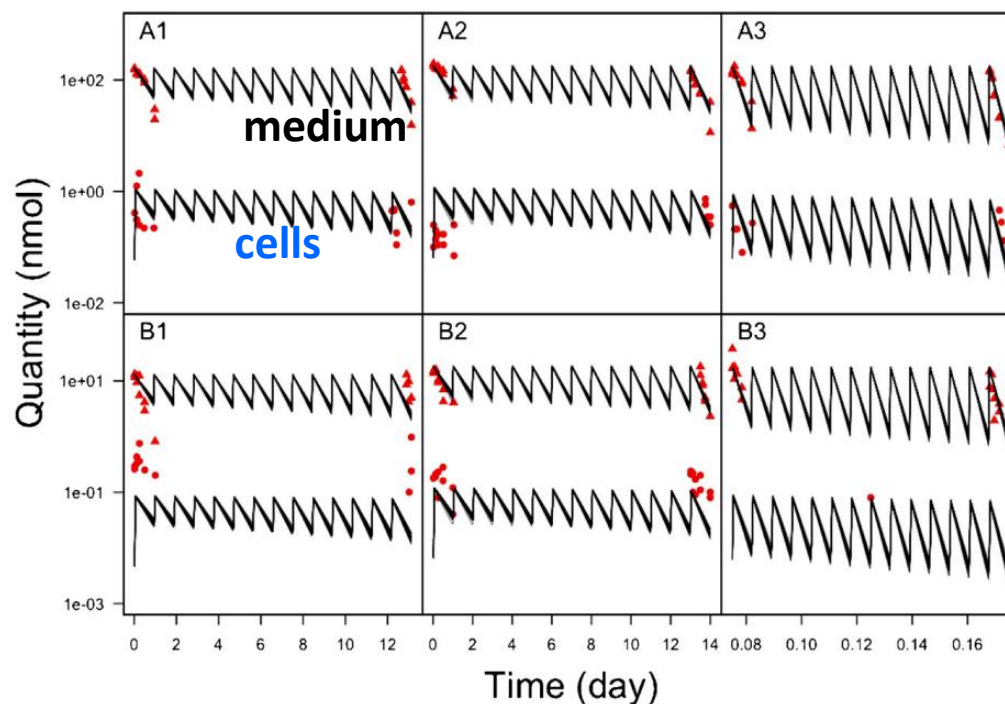
# Biokinetics “FEASIBLE” after Repeated dosing

Hydrophilic  
substance:

$\log P_{O/W} =$   
2.23;

a nonprotein-  
bound  
fraction of  
0.01

Repeated  
dosing  
→



*Truissi et al., Toxicology Letters 233 (2015) 172–186*

*Mielke, et al. Arch Toxicol, 91 (2017), 1663-1670.*

# Protein binding: protein content in the media → key for the dose/concentration causing a toxic effect



## QIVIVE : Quantitative In vitro to in vivo extrapolation Case-study 1

- Modelling the in vivo doses starting from the in vitro actual measured concentrations in the medium and cells;
- Conditions used in the in vitro study (protein-free in vitro culture medium) : Set the protein binding at 0% for the modelling;
- ≠ conditions → in vitro: the culture medium could not bind the substance, in vivo the substance may have high affinity for plasma proteins;
- Higher in vitro free fraction than in the blood in vivo → ≠ organ/blood partition coefficient.

### Protein binding

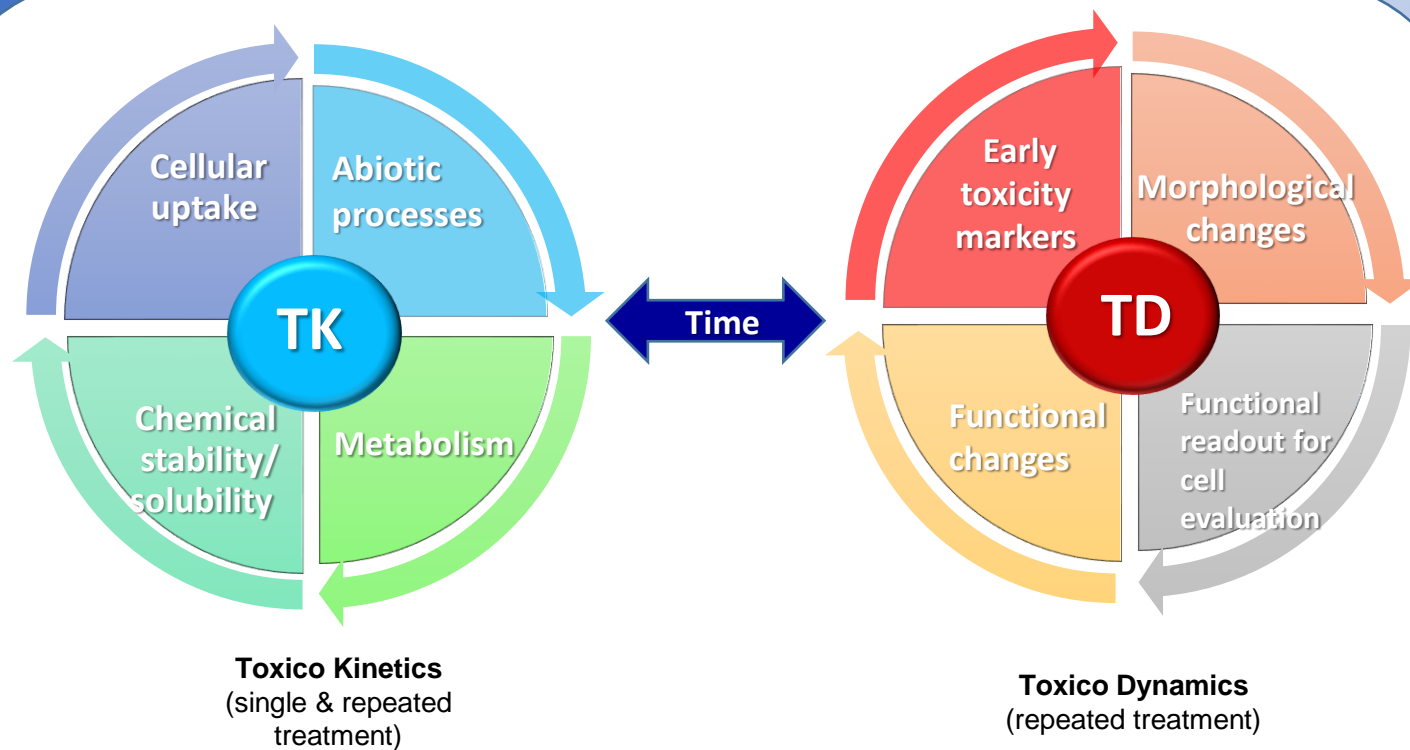


In vitro conditions must be considered and compared to the in vivo situation, particularly, for protein binding.

*Algharably, Di Consiglio et al., (2022, in press)*

*In Vitro–In Vivo Extrapolation by PBPK modeling: Experience With Three Case Studies and Lessons Learned. Front.Toxicol. 4:885843.doi: 10.3389/ftox.2022.88584*

# Kinetics vs Dynamics



*Modified from Di Consiglio et al., Reproductive Toxicology 98 (2020)*



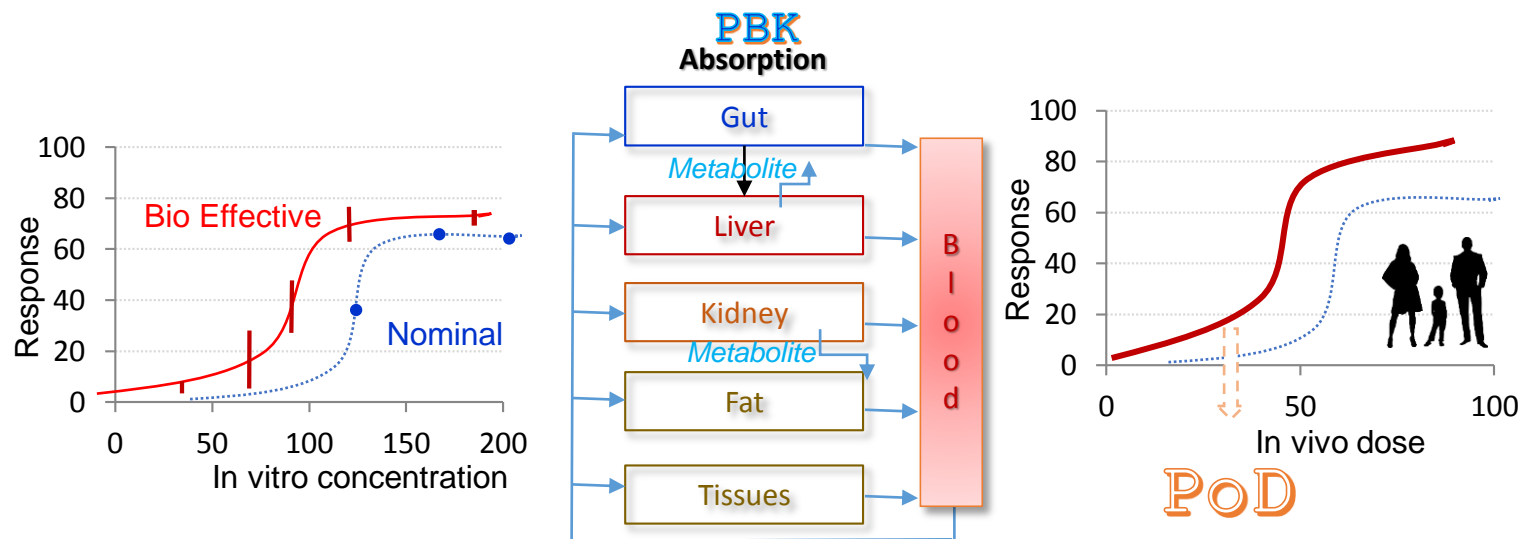
# Nominal vs actual concentrations

QIVIVE for the relevant adverse effects Case-study 2

In vitro measured intracellular concentration →

**Bio Effective Concentration** → Tissue/organ concentration

Appropriate  
QIVIVE



Algharably, Di Consiglio et al., (2022, in press)

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# Nominal vs actual concentrations

QIVIVE for the relevant adverse effects Case-study 2

In vitro measured intracellular concentration →

**Bio Effective Concentration** → Tissue/organ concentration

← **Appropriate QIVIVE**

**PBK**  
Absorption

Gut

100

QIVIVE dose (mg)

Doses in patients with  
neurological side effects (mg)

BMDU based on measured in  
vitro intracellular AUC<sub>0-24</sub>

593

BMDU based on nominal  
concentration

10833.3

400–500

0 50 100 150 200  
In vitro concentra

In vivo dose

**Underestimation of potency**

**PoD**

**IMP.** to estimate the actual PoD:  
Phys-Chem properties; kinetic parameters  
e.g. protein binding; metabolism

Algharably, Di Consiglio et al., (2022, in press)

In Vitro–In Vivo Extrapolation by PBPK modeling: Experience With Three Case Studies and  
Lessons Learned. *Front.Toxicol.* 4:885843.doi: 10.3389/ftox.2022.88584

# Predictive models: Improving cell models



**Complex cell models:** e.g. *human* induced pluripotent stem cells (hiPSCs) and their differentiated derivatives, cultured in *3D* to improve the level of physiological complexity; *organoids* in *microfluidic devices*; engineered tissues or *multiorgan systems*

**In vitro assays** should be associated to  
**HUMAN relevant** key events,  
in **HUMAN relevant** cellular models

Metabolic competence and transporter  
expression of the cell models →  
relevance to the human in vivo situation

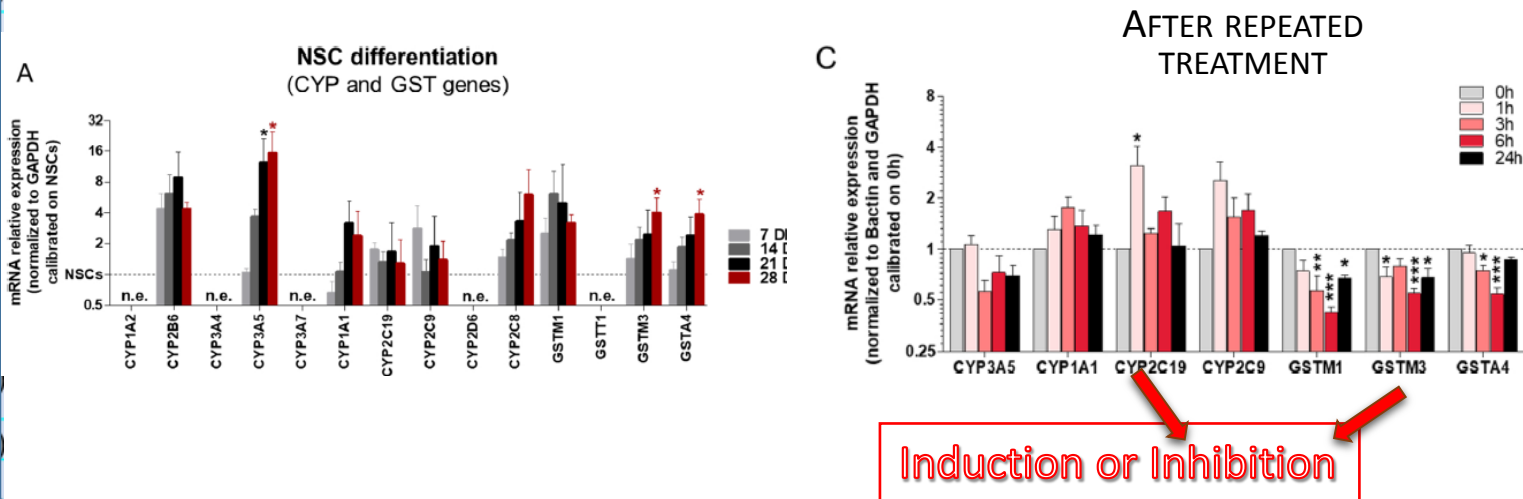
Metabolic  
capacity  
Transporter  
expression

# Importance to consider the BIOTRANSFORMATION and TRANSPORT

Biotransformation and transport capacity of the test system used.

Case-study 3

- Are the elements of concern expressed?
- Is it possible to predict systemic exposure to both the parent molecule and relevant metabolites for a correct NGRA?
- Are the right test substances (i.e. and metabolite) tested in vitro? Is it possible to calculate in vitro clearance to be extrapolated to the in vivo



**IMP.** to focus the risk assessment on both the parental substance and its “MAJOR” metabolite(s) → clearance calculation + time component



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The case of systemic exposure to  
phenoxyethanol and  
its major metabolite phenoxyacetic acid in  
body lotions

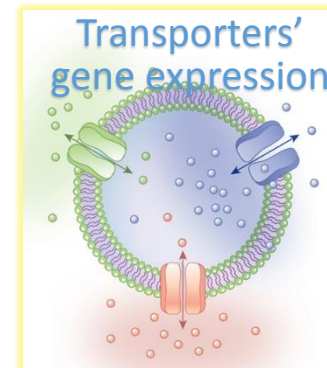
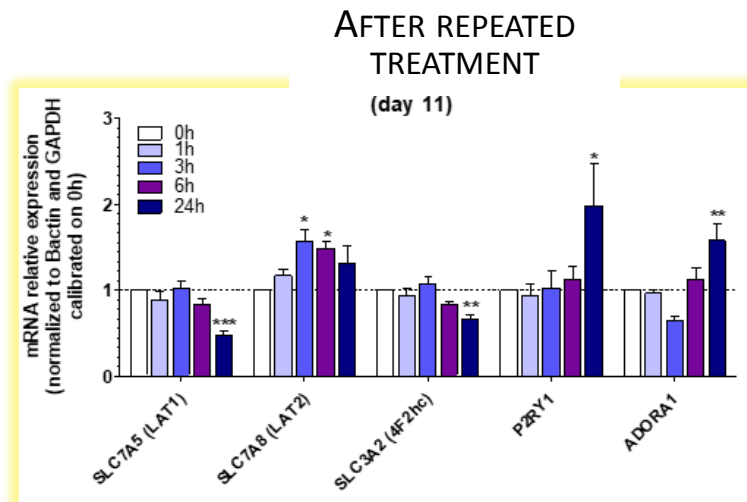


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# Predictive models: Improving cell models



**Organ-on-Chip (OoC)**: among new non-animal in vitro technologies → *considerable interest* within the scientific community:

They recreate *body physiology capacities* to transform science, in the field of biomedical research, drug development, consumers safety and personalised medicine.

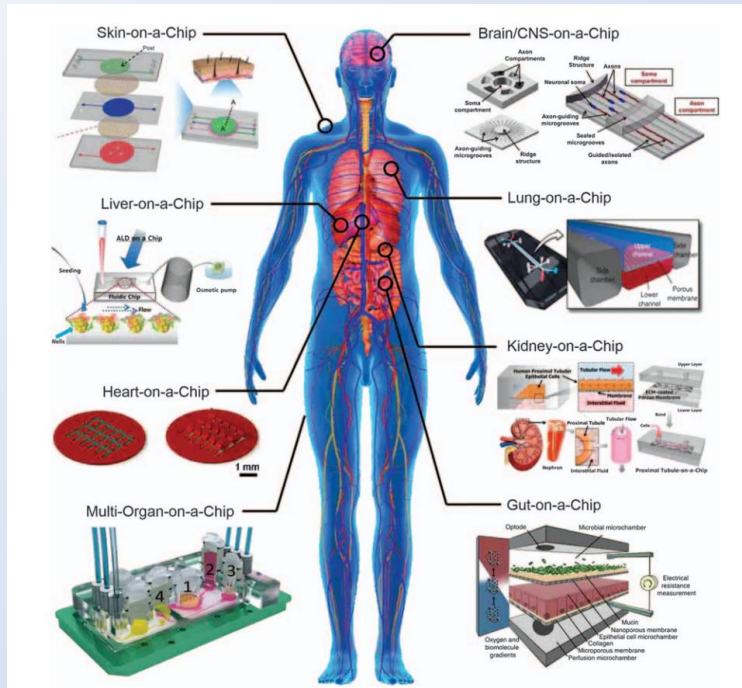
*Full qualification* process → necessary to demonstrate the relevance, promoting implementation both by end users and regulators.

***Qualification*** for contexts instead of a full validation: in order to support decision-making in regulatory setting.



# Organ-on-chip (OoC)

ongoing /



Microfluidic microphysiological systems (MPS)



EURL ECVAM

Piergiorgio et al., Lab Chip, 2021



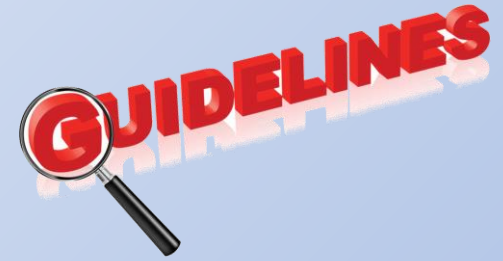
Analysed the state of the art of standards in OoC, describing the technical and biological aspects of OoC, focusing on standardisation needs and opportunities.

- **Context of use**: circumstances under which the MPS is applicable + limitations (e.g. applicability of the test method for a particular class of compounds);
- **Stepwise approach** (start simple...) taking into account both safety and efficacy testing;
- **Reliability**; reproducibly over time;
- **Relevance**, how the test method correctly measures or predicts the biological effect;
- **Qualification** implies the availability of defined test methodology including aspects related to in vitro to in vivo extrapolation (**QIVIVE**), e.g. **biokinetics**;
- **Standardization**: to allow the regulatory acceptance of OoC.

**P**reliminary  
**A**ssessment  
**R**egulatory  
**R**elevance



# Standardization



Universal approach to validation or harmonization :  
probably unrealistic (too long, might not be possible) →  
application of OoC devices in the short/medium term  
primarily *target specific* and *well-defined contexts of use*.

**Standardization** by verifying:

- ✓ *reproducibility* and *robustness* of results intra- and inter-lab;
- ✓ *consistency* or *compatibility* of cell/tissue types and sources;
- ✓ *producer compatibility* among chips or modules  
also in interconnection into a multi-organ system.

***Standardization will aid in the concrete incorporation of OoC-based studies into regulatory workflows and decision-making contexts:  
Ex. EURL ECVAM, liver and brain 3D models for DNT applications  
→ reliability and relevance of OoC for its intended purpose***

# Regulatory requirements in NGRA (NAMs+Biokinetics)



*\*Take  
home message*

- For scientific acceptance agreed standards needed for using tools, recording methods or protocols, data analysis and reporting → transparency & consistency;
- Formal validation process not possible?  
Application of new approaches in an accepted fit-for-purpose context.
- **General challenges of in vitro systems & study design:**
  - a. Key biological and fisiological functions of organ to be represented
  - b. Stability of the model over time
  - c. Does the in vitro system have the adequate metabolic/transporter capability?
  - d. What is/are the relevant substance(s) (parental or metabolites) and at which internal exposure conditions?
  - e. How does the chemical behave in the in vitro system with respect to protein, lipid and plastic binding and evaporation?

# Regulatory requirements in NGRA (NAMs+Biokinetics)



*\*Take  
home message*

- QIVIVE:
  - a. What are the internal dose of a chemical that are reached in a certain exposure scenario ?
  - b. How do these doses relate to in vitro bioeffective concentrations ?
  - c. Extrapolation of an in vitro biological effect concentration (e.g., BMC10) to an equivalent oral/skin/inhalation dose.

# Thank You For Your Attention!



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National Representative

Network PARERE Italiano



**Special thanks to all the colleagues, who took part to the cited papers and made this dissertation possible.**

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